The Effects of a Probiotic (*Bacillus clausii*) in Acute Kidney Injury in a Rat Model of LPS-Induced Endotoxemia

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ABSTRACT

Objective: This study aims to examine the role of a probiotic bacterium, *Bacillus clausii*, on oxidative stress in a lipopolysaccharide (LPS)-induced acute kidney injury (AKI) rat model.

Materials and Methods: The rats were divided into four groups: Control group, LPS group (1.5 mg/kg, LPS), Probiotic+LPS group in which LPS was given after administration of *Bacillus clausii* as a probiotic for 21 days, and Probiotic group. The kidneys of the rats were removed 24 hours after LPS injection. Total antioxidant status, total oxidant status, oxidative stress index, malondialdehyde and myeloperoxidase (MPO) values were biochemically determined in the kidneys. Furthermore, the kidney tissue samples were immunohistochemically stained for interleukin-6 (IL-6) and tumor necrosis factor (TNF)-α expression, and leukocyte distribution.

Results: Endotoxemia caused an increase in oxidative stress (p<0.001), lipid peroxidation (p<0.01), MPO activity (p<0.001), and the expression of IL-6 (p<0.001) and TNF-α (p<0.001). The administration of probiotic ameliorated oxidative stress, lipid peroxidation, and myeloperoxidase activity, and resulted in decreased IL-6 and TNF-α reactions that were elevated with LPS treatment.

Conclusion: The results suggest that *Bacillus clausii* as a probiotic bacterium may have an antioxidative property in LPS-induced AKI.

Keywords: Probiotic, acute kidney injury, endotoxemia, oxidative stress, *Bacillus clausii*

INTRODUCTION

Microbiota consists of various microorganisms (bacteria, fungi, and protozoa) (1). There is a symbiotic and mutualistic relationship between microorganisms in the gut microbiota and host (2). In mammals, the microbiota is composed mainly of four main phyla of eubacteria: Bacteroidetes, Firmicutes, Actinobacteria, and Proteobacteria (3, 4). The microbiota plays significant roles in various physiological events, such as protection against infection, recovery from disease, drug metabolism, nutritional status, and vitamin synthesis (3, 5).

The change in gut microbiota is called dysbiosis, leading to a disrupted interaction between host and microbes. The resulting systemic complications and disease may follow deterioration in microbiota homeostasis caused by excessive increase or depletion of specific bacterial species (6). Dysbiosis relates to several diseases such as obesity, diabetes, and inflammatory bowel and critical disease (7, 8). It was shown that the loss of “health promoting” bacteria and dysbiosis could contribute to infections, organ failure, and sepsis in the intensive care unit (ICU) (7). Some members of the gut microbiota generate anti-inflammatory molecules and proteins, while the microbiota can cause inflammatory responses. It is thought that the loss of useful microorganisms can disrupt nutrition in humans and trigger inflammatory responses. Hence, modification of the
microbiota composition can restore nutritional deficiencies and anti-inflammatory effects (9).

Probiotics have attracted attention as an opportunity to treat and prevent different diseases such as diabetes, obesity, allergies, and infections (10-12). In recent years, the question has been raised as to whether restoration of ICU patients through probiotics or symbiotics to interfere with the preservation and degradation of the microbiota would be an optimal interference to prevent infection and improve recovery.

The supplementation of probiotics offers a significant approach to critical illness therapy because of the important roles of microbiota in the modulation of nutrient production and absorption, and inflammation (9). Probiotics are live microorganisms which, when they are administered in adequate amounts, confer a health benefit on the host (13).

Different dietary supplements, functional foods, or probiotic strains that can be beneficial for human health have been designed. Data obtained from studies have supported the use of bacterial spore formers as probiotics and food supplements (14).

Bacillus spp., one of the spore-forming probiotics, are Gram-positive, aerobic or facultative anaerobic bacteria, and can survive in heat, acidic stomach, and other extreme environmental conditions (15-17). Therefore, spores of Bacillus spp. are generally used as probiotic preparations for different purposes (18).

Acute kidney injury (AKI) is a serious and common complication of sepsis in ICU patients (19). The pathophysiology of AKI in endotoxemic shock or sepsis is complex, and contains hemodynamic changes, inflammation, and damage. Moreover, sepsis-induced immune responses include the activation of both pro-inflammatory and anti-inflammatory events (20, 21). In the literature, it has been shown that the gut microbiota plays a role in kidney disease (22-25). Although the function of gut microbiota has been determined in kidney disease (22-25), the relationship between AKI and microbiota is not yet fully understood.

This study aimed to investigate the effects of the probiotic bacterium Bacillus clausii on renal oxidative stress in a lipopolysaccharide (LPS)-induced AKI model.

MATERIALS AND METHODS

Animals and Experimental Protocol

The experiments were designed with 27 male Wistar albino rats (3 months old, 300-400 g) supplied from Bezmiâlem Vakif University, Istanbul, Turkey. The animals were allowed free access to a pelleted diet and tap water. The study was approved and reviewed by Bezmiâlem Vakif University Animal Experiments Local Ethics Committee (Decision no: 2021/54).

The rats were divided into four groups. The control group (n=6) was injected intraperitoneally (ip) with 0.9% NaCl solution (physiological saline) after physiological saline was given by gavage for 21 days. In the LPS group (n=7), LPS (1.5 mg/kg LPS, ip, E. coli, Serotip 0111: B4, Sigma, Missouri, USA) was injected ip after physiological saline was given by gavage for 21 days. In the Probiotic+LPS group (n=6), LPS was injected ip into animals after the administration of B. clausii (1.25 ml, 1×10⁵ CFU per animal via gavage) as a commercial probiotic (Enterogermina®, Sanofi, Turkey) for 21 days. Probiotic group (n=8) was ip treated with physiological saline after B. clausii was given by gavage for 21 days. All applications were done at the same time every day.

The rats were anesthetized 24 hours after LPS injection. The kidney samples were removed for biochemical and immunohistochemical analyses.

Biochemical Analyses

The tissue samples were homogenized and centrifuged at 3000 rpm for 20 minutes at +4 ºC (Sorvall Super T21, Benchtop Centrifuge). The supernatants were collected from homogenates.

Total oxidant status (TOS) (µmol H₂O₂ eq/L), total antioxidant status (TAS) (mmol Trolox eq/L), and oxidative stress index (OSI) (arbitrary unit) were measured in the kidney tissues using commercial kits (Rel Assay Diagnostics; Gaziantep, Turkey).

Malondialdehyde (MDA) in the kidney was measured using the spectrophotometric method (26). The results were expressed as nmol/mg protein.

Myeloperoxidase (MPO) activity was measured according to the method of Krawisz et al. (1984) (27). The results were expressed as µmol/mL.

Protein levels in homogenate/postmitochondrial fractions were measured using the bicinchoninic acid reaction (28).

Immunohistochemical Analyses

The kidney samples were fixed in a neutral buffer formaldehyde solution, and histologically processed as previously described (29). The kidney sections (5 µm thick) were stained with tumor necrosis factor (TNF)-α (diluted 1/200) (Abcam ab66579), interleukin-6 (IL-6) (diluted 1/200) (Abcam, 6672), and MPO (diluted 1/75) (Thermo Fisher Scientific, RB-373-A) antibodies. The antibody staining was scored as previously described by Legrand et al. (29).

Statistical Analysis

Significant differences between groups were estimated using one-way ANOVA with Tukey’s post-test using GraphPad Prism version 5.0 for Windows (GraphPad Software, San Diego, CA, USA). A value p<0.05 was considered statistically significant. The results were expressed as the mean±SEM.

RESULTS

Biochemical Results

TOS levels increased in renal tissues of LPS-injected animals (p<0.001), and diminished in the Probiotic+LPS group, but was high with respect to the control (p<0.001). In the Probiotic group, TOS levels elevated according to the control (p<0.05).
TAS levels reduced in renal tissues of the LPS group. Administration of the probiotic caused an increase in TAS levels (p<0.01) in the Probiotic+LPS group. Moreover, TAS level was similar to the control group. TAS levels increased in the Probiotic group compared with the control group (p<0.001) (Figure 1B). OSI values increased (p<0.001) in the LPS group compared with the control group. In the Probiotic+LPS group, OSI values decreased compared with the LPS group (p<0.05). OSI values of probiotic treated groups were similar to the control (Figure 1C).

MDA levels increased in LPS-treated animals (p<0.01) and reduced in the Probiotic+LPS group (p<0.05) when compared to this group. MDA levels of the Probiotic+LPS and Probiotic groups resembled the control group (Figure 2A). MPO activity elevated in the LPS group (p<0.001) and diminished in the Probiotic+LPS group when compared to this group. In the Probiotic+LPS group, it was high with respect to the control group (p<0.01). MPO activity of the Probiotic group was higher than the control group, but it was not statistically significant (Figure 2B).

**Immunohistochemical Results**

IL-6 and TNF-α reaction and MPO-stained leukocytes are presented in Figure 3. LPS-induced endotoxemia caused an increase in TNF-α and IL-6 reactions in the kidney tissues (p<0.001). The probiotic administration decreased IL-6 (p<0.001) and TNF-α reaction (p<0.01) compared with the LPS group. The distribution of MPO-stained leukocytes was marked in both glomeruli and peritubular areas in the LPS groups (p<0.001) in comparison to the control group. The probiotic did not affect the distribution of MPO-stained leukocytes in the kidney tissue in LPS given animals (Figure 3).

**DISCUSSION**

AKI is an important health problem (19). Sepsis, endotoxemia, ischemia reperfusion, or nephrotoxins cause AKI (30). The fundamental interaction between kidney and gut microbiota is an important regulating factor in AKI (22, 23). Gut microbes are associated with physiological and pathophysiological processes...
in the kidneys (23). Short chain fatty acids (SCFAs) that gut microbes produce improved sepsis-induced AKI (24) and reduced kidney damage and levels of reactive oxygen species and cytokines after kidney ischemia-reperfusion injury (25). Renal structural injury and functional decline following ischemia-reperfusion injury were more severe in germ-free mice compared with control mice (31). It was reported that severely abnormal gut microbiota in patients with renal failure led to aggravate kidney disease (32). Protecting the balance of the gut microbiota was stated to be important for ameliorating dysbiosis that results in immunological dysfunction, inflammation, and renal disease (23). Although the function of gut microbiota has been determined in kidney disease, the link between AKI and microbiota is not yet fully understood. Major advances in studies of the composition and functions of gut microbiota have opened up the opportunity for application of treatment strategies for AKI patients (22).

Changes in gut microbiota control metabolic endotoxemia characterized by translocation of lipopolysaccharides in systemic circulation, and induction of oxidative stress and inflammation in obesity and diabetes (12). Ávila et al. suggest that probiotics and fecal microbiota transplantation could present immunomodulatory advances and may be functional for treatment of pediatric septic patients (33). The different probiotics could reduce the oxidative damage and inflammation in intestines of animals with sepsis (33). Probiotics prevent intestinal and liver damage in sepsis induced with administration of LPS and D-galactosamine (11). The selected probiotic isolates reduced the LPS-caused inflammation in murine macrophage cell line (34). Lactobacillus salivarius reduced inflammation and oxidative stress in cisplatin-induced AKI. These effects were partially mediated by modulating the gut environment (35).

In a rat study of sepsis that was induced by injection of LPS after the administration of Lactobacillus rhamnosus gg as probiotic (1x10^7 CFU/day) for 10 days, the probiotic was revealed to have exerted protective effects (36). Similarly, LPS induced oxidative stress and inflammation in the present study. Moreover, the pre-treatment with a probiotic prevented oxidative stress, and exerted protective effects in endotoxemia that was induced by LPS. The probiotic administration after LPS treatment induced a humoral immune response by increasing the levels of immunoglobulins E, A, G, and M (37).

A number of Bacillus strains as probiotic have been studied for their potential functions in vivo and in vitro models (38, 39). B. clausii is an endospore-forming, gram-positive, facultative alkaliphilic rod bacterium (15), and is able to grow under aerobic and anaerobic conditions and survive gastric acidity (16, 17). The role of the Bacillus species ranges from the probiotic nature

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Figure 3. The immunoreactivity and distribution of tumor necrosis factor (TNF)-α, interleukin-6 (IL-6), and myeloperoxidase (MPO)-stained leukocytes (→) in kidney sections, and HSCORE values of immunoreactive reactions in the experimental groups. Bar: 50 μm. LPS: lipopolysaccharide. ***p<0.001 vs. Control group. **p<0.01, *p<0.001 vs. LPS group.
of *B. clausii*, *B. subtilis*, *B. pumilus*, *B. coagulans* and other strains, to biological control agents (*B. sphaericus* and *B. thuringiensis*) and pathogenicity (*B. cereus* and *B. anthracis*) (18). The effects of *B. clausii* have been associated with antimicrobial and immunomodulatory properties (40). *B. clausii* had a role in production of pro-inflammatory and anti-inflammatory cytokines (41). Furthermore, *B. clausii* strains release antimicrobial substances (40). The probiotic *B. clausii* strain inhibits the cytotoxic effects caused by *B. cereus* and *Clostridium difficile* toxins (38). *B. clausii* may reveal immunomodulating activity in nasal lavage of allergic children (10). *B. clausii* inhibited secretion of pro-inflammatory cytokines and reactive oxygen species production in rotavirus-infected cells (39). It was demonstrated that pretreatment with probiotic capsules of *Bacillus* species spores caused a significant decrement in pro-inflammatory cytokines (42). Treatment with *B. clausii* UBBC07 importantly attenuated oxidative stress by increasing catalase and SOD, and decreasing MDA in acetoniphen-induced uremia in rats (43). In our study, the treatment with *B. clausii* importantly ameliorated the oxidative stress and lipid peroxidation caused by LPS injection.

Current data showed that Enterogermina, a spore-based probiotic formulation of *Bacillus* spp., is a trusted probiotic to manage acute diarrhea and intestinal infections (44) and to decrease anti-*Helicobacter pylori* treatment (45, 46), and exhibits immunomodulatory and antimicrobial activities (10, 40).

In the study, the effects of probiotic *B. clausii*, marketed as Enterogermina®, on rat kidneys were examined in LPS-induced AKI. Similar to current data, the present study indicated that probiotic *B. clausii* has antioxidant and anti-inflammatory properties. The results revealed that LPS gave rise to increase TOS and decrease TAS. It can be said that LPS-induced endotoxemia caused oxidative stress as indicated with an increase in the oxidative stress index. Furthermore, it was found that endotoxemia increased the levels of lipid peroxidation and inflammation in the kidney tissue. In addition, expression of pro-inflammatory cytokines such as TNF-α and IL-6 and leukocytes infiltration was determined in kidney tissues of endotoxemic rats. The pre-treatment with *B. clausii* ameliorated the oxidative stress, lipid peroxidation, and inflammation, and decreased expression of pro-inflammatory cytokines in LPS-induced endotoxemic rats. However, probiotic treatment did not affect the distribution of leukocytes in kidney tissues.

**CONCLUSION**

Our results suggested that probiotic *B. clausii* may have an antioxidative and anti-inflammatory property in LPS-induced AKI. Therefore, the present study supports the efficacy of *B. clausii* in the attenuation of LPS-induced AKI. Nevertheless, further studies should be performed to determine the exact role and effect mechanisms of this probiotic in kidney disease.

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**REFERENCES**

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